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DATE: Tuesday, February 13, 2007

Hide?	Set Name	Query	Hit Count
	DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI,TDBD; PLUR=YES; OP=OR		
☐	L23	9623899	5
☐	L22	L21 and facs	1
☐	L21	9638553	5
☐	L20	9504824 and (light scattering)	1
☐	L19	9504824 and facscan	0
☐	L18	9504824 and facs	0
☐	L17	9504824	10
	DB=USPT; PLUR=YES; OP=OR		
☐	L16	9504824	0
☐	L15	l7 and p21	3
☐	L14	5612185 and p21	0
☐	L13	5612185 and (retroviral library)	3
☐	L12	5612185 and (six parameter)	6
☐	L11	L9 and (six parameter)	64
☐	L10	L9 and (library retroviral)	33
☐	L9	L8 and (cellular phenotypes)	70
☐	L8	L7	80
	DB=PGPB,USPT,USOC,EPAB,JPAB,DWPI,TDBD; PLUR=YES; OP=OR		
☐	L7	L6 and multiparameter	158
☐	L6	facscan	5075
☐	L5	9727212	6
☐	L4	L3 and (retroviral library)	3
☐	L3	L1 and (FACS)	9
☐	L2	L1 and (FACS multiparameter)	10
☐	L1	6365362	34

END OF SEARCH HISTORY

(FILE 'HOME' ENTERED AT 11:22:41 ON 13 FEB 2007)

FILE 'CA' ENTERED AT 11:23:11 ON 13 FEB 2007

L1	5530	S	FACS
L2	1462	S	L1 AND FLOW CYTOMET?
L3	45	S	L1 AND LIGHT(2W)SCAT?
L4	1	S	L3 AND FLUOR?(3W)(UPTAKE OR RELEASE)
L5	5529	S	L1 NOT L4
L6	0	S	L5 AND FIVE(3W)PARAMETERS
L7	165	S	L5 AND ANNEXIN
L8	60	S	L7 AND FLUORES?
L9	0	S	L8 AND ENZYME(2W)ACTI?
L10	4	S	L8 AND ENZYME
L11	56	S	L8 NOT L10
L12	7	S	L11 NOT 1999-2007/PY
L13	49	S	L11 NOT L12
L14	1449	S	L2 NOT L13
L15	16	S	L14 AND MULTIPARAMETER

ANSWER 16 OF 16 CA COPYRIGHT 2007 ACS on STN

TI C-kit is expressed by primitive human hematopoietic cells that give rise to colony-forming cells in stroma-dependent or cytokine-supplemented culture

AU Simmons, Paul J.; Aylett, Gabriella W.; Niutta, Silvana; To, L Bik; Juttner, Christopher A.; Ashman, Leonie K.

SO Experimental Hematology (New York, NY, United States) (1994), 22(2), 157-65
CODEN: EXHMA6; ISSN: 0301-472X

PY 1994

AB Using monoclonal antibody (MAB) YB5.B8, the authors have examined the expression of the c-kit protein, the receptor for the hematopoietic cytokine stem cell factor (SCF), on primitive hematopoietic cells. Bone marrow mononuclear cells (BMMNC) enriched for immature cells by differential agglutination using the lectin soybean agglutinin (SBA) were subjected to multiparameter fluorescence activated cell sorting (FACS) based on light-scattering properties, the expression of the c-kit protein and the CD34 antigen, and the retention of the vital fluorescent dye, Rhodamine 123 (Rh123). Sorted populations were assayed for their content of directly clonogenic progenitor cells (colony-forming units-granulocyte/macrophage [CFU-GM], burst-forming units-erythroid [BFU-E], and multipotential colony-forming units [CFU-Mix]) and for the presence of more primitive progenitor cells (pre-CFU). The latter were assayed by their ability to initiate and sustain hematopoiesis in a standard stromal cell-dependent culture system and their capacity for de novo generation of clonogenic progenitors in response to a combination of 6 recombinant hematopoietic cytokines in a stroma-independent suspension culture assay. A mean of 76% of CD34+ cells were found to coexpress c-kit. The majority of directly clonogenic cells (98% of CFU-GM, 98% of CFU-Mix, and 85% of BFU-E) were found in the CD34+ c-kit+ fraction. Similarly, all pre-CFU were recovered in the CD34+c-kit+Rh123dull fraction, irresp. of whether the cells were maintained on marrow stromal cells or in cytokine-supplemented liquid culture. A mean of 87% of the CD34+Rh123dull cells also expressed c-kit. Since SCF has been reported to act as a growth factor for early lymphoid cells as well as myeloid cells, the authors looked for coexpression of c-kit and early lymphoid markers in the CD34+ population by multiparameter flow cytometry. Coexpression of c-kit on a minority of cells with markers of B or T lineages was observed The majority of early lymphoid cells, however, appeared to lack c-kit expression. This was confirmed by the finding that only 4% of c-kit+CD34+ cells showed terminal deoxynucleotidyl transferase activity, compared with 25% of the c-kit-CD34+ cells.